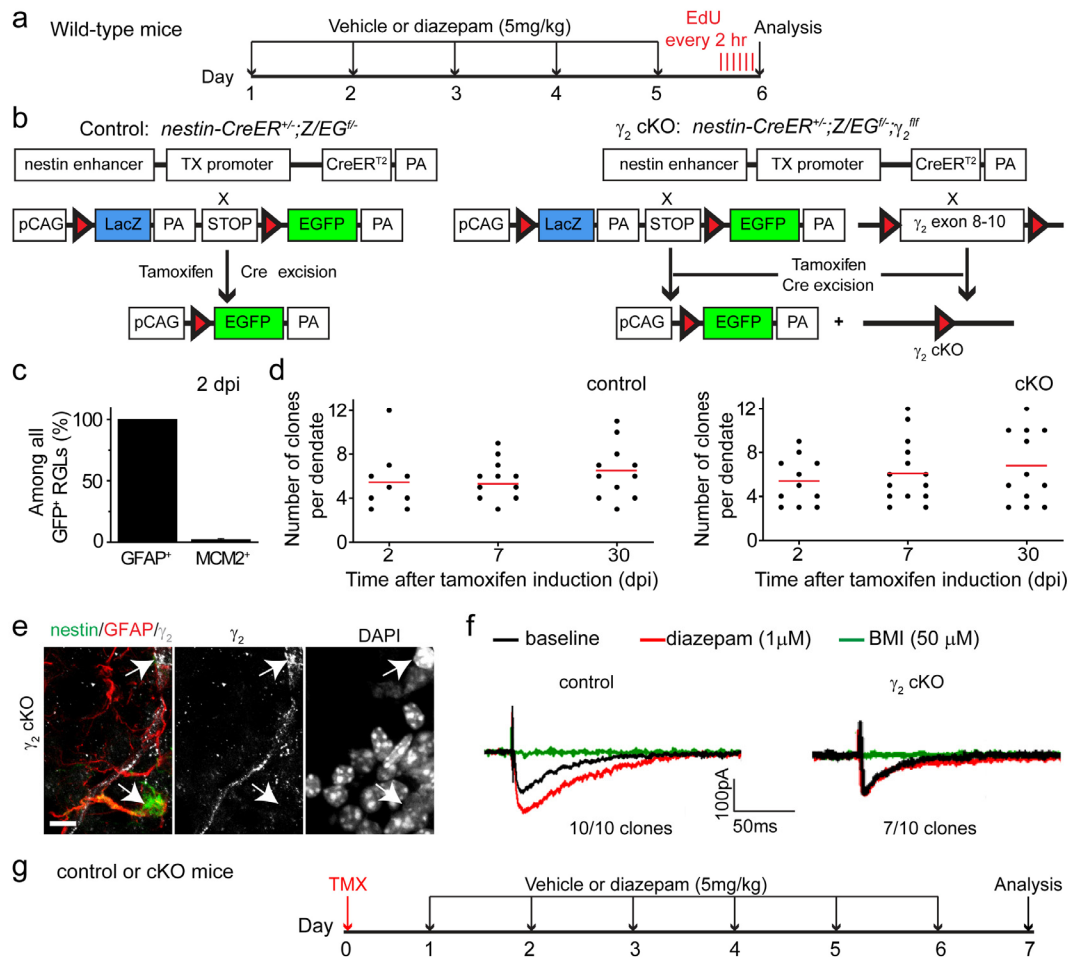
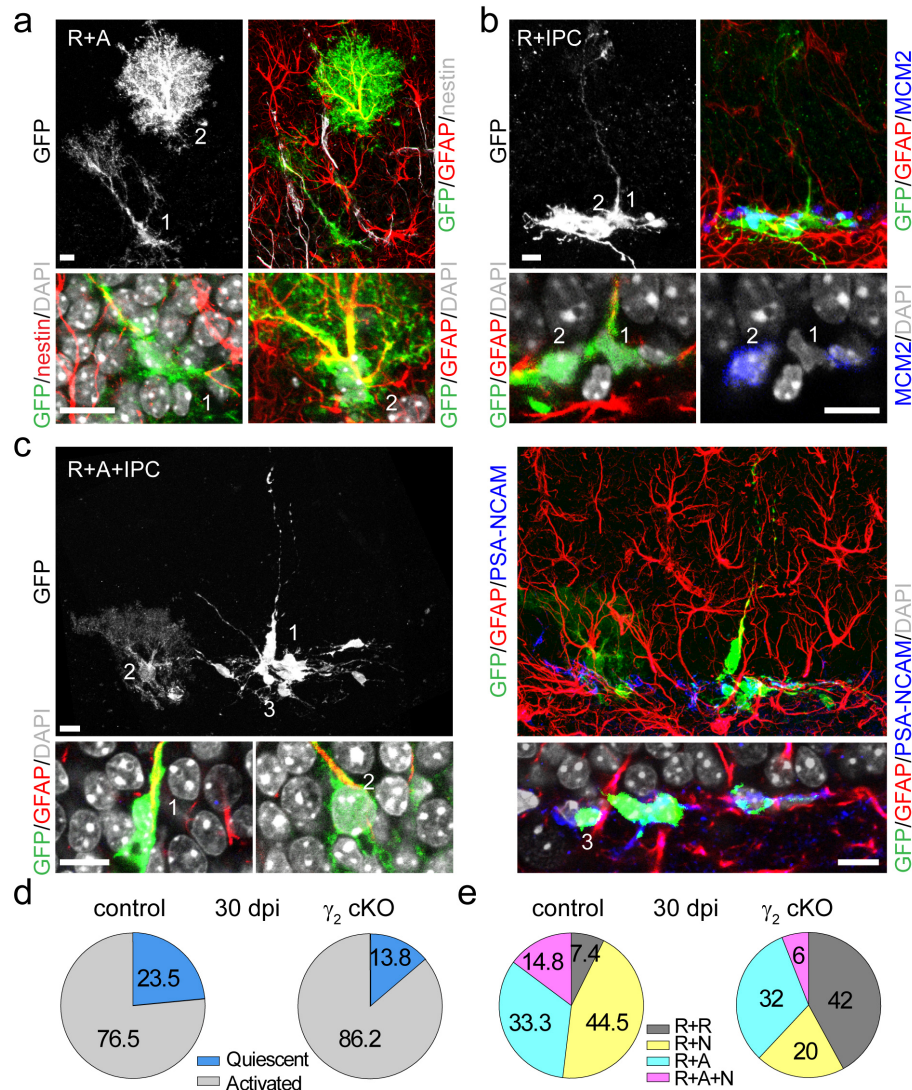


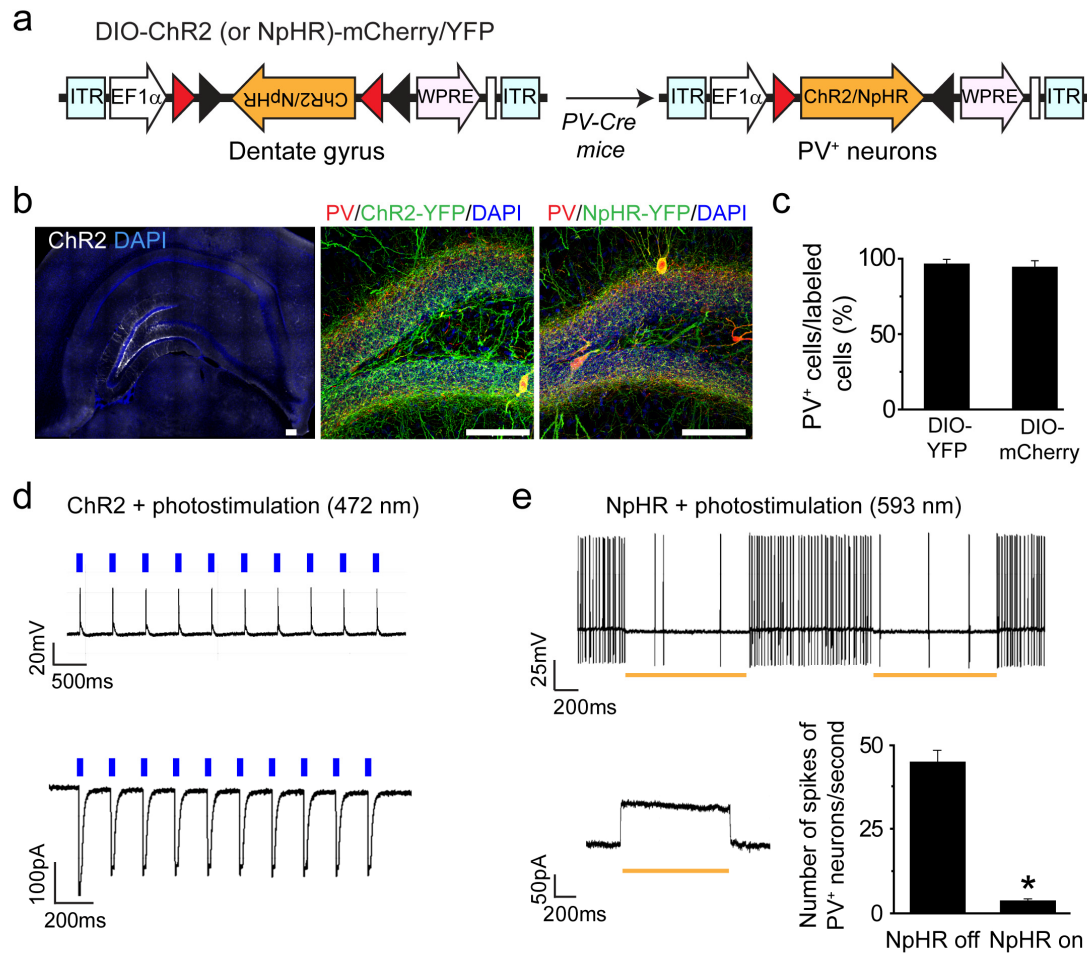
**Supplementary Figure 1. Basic characterization of GFP<sup>+</sup> RGLs in the dentate gyrus of adult *nestin-GFP* mice.** **a**, Sample confocal images of GFP, DAPI and immunostaining of GFAP and MCM2 in the dentate gyrus of adult *nestin-GFP* mice. Scale bar: 10  $\mu$ m. **b**, Quantification of the percentages of GFAP<sup>+</sup> or MCM2<sup>+</sup> RGLs among all GFP<sup>+</sup> RGLs in the dentate gyrus of adult *nestin-GFP* mice. Values represent mean  $\pm$  s.e.m. ( $n = 3$  animals). **c-h**, Sample traces of whole-cell voltage-clamp recording from GFP<sup>+</sup> RGLs in slices acutely prepared from adult *nestin-GFP* mice ( $V_m = -65$  mV). Shown are sample traces of a GFP<sup>+</sup> RGL in response to puffs of GABA (200 mM), in the presence or absence of diazepam (1  $\mu$ M) or bicuculline (BMI; 50  $\mu$ M, **c**), and response to muscimol (200 mM, **d**). Also shown is an imaging of a recorded cell loaded with dye through the recording pipette (**c**). Shown in (**e**) are sample confocal images of GFP<sup>+</sup> cells (green) immunostained for GFAP (red) and  $\gamma_2$  (white) in the dentate gyrus of adult *nestin-GFP* mice. Scale bars: 10  $\mu$ m (left) and 5  $\mu$ m (right for insert). Shown in (**f**) are traces indicating a lack of evoked postsynaptic currents (PSCs) in a GFP<sup>+</sup> RGL in response to the field stimulation of the dentate granule cell layer (top) and no spontaneous synaptic currents (SSCs; bottom). Shown in (**g**) are sample traces of a GFP<sup>+</sup> RGL in response to a train of field stimuli (8 Hz, 100 stimuli) of the dentate granule cell layer, in the presence or absence of BMI (50  $\mu$ M). Note tonic responses induced by the stimuli. Shown in (**h**) are sample traces from a GFP<sup>+</sup> RGL in response to bath application of bicuculline (BMI; 100  $\mu$ M; top), revealing the basal level of tonic responses, and from a GFP<sup>+</sup> RGL in response to bath application of sucrose (30 mM) to increase the presynaptic release probability, followed by BMI treatment (bottom), demonstrating the presence of a tonic response to ambient GABA, but not synaptic GABAergic responses.



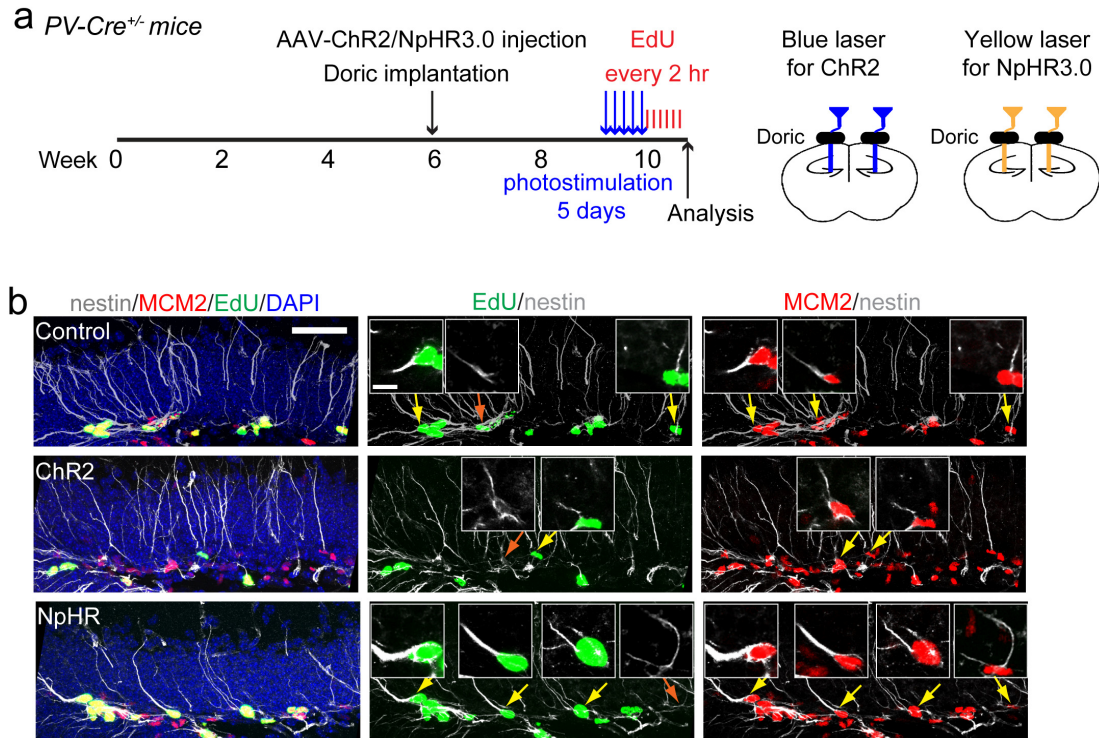
**Supplementary Figure 2. Analysis of the role of  $\gamma_2$ -containing GABA<sub>A</sub>Rs in regulating RGLs in the adult dentate gyrus.** **a**, Experimental scheme for diazepam experiment (related to Fig. 2a-b). Wild-type adult mice were *i.p.* injected with diazepam (5 mg/kg body weight; once daily for 5 days), followed by 6 injections of EdU (41.1 mg/kg body weight) at 2 hrs apart, and then processed for immunohistology 2 hrs after the last EdU injection. **b**, A schematic diagram for *in vivo* clonal analysis of individual RGLs with or without conditional  $\gamma_2$  deletion. **c-d**, Basic characterization of the clonal analysis approach. Shown in (c) is the quantification of immunohistological characterization of GFP<sup>+</sup> RGLs in the subgranular zone of the adult dentate gyrus of control mice at 2 dpi. Values represent mean  $\pm$  s.e.m (n = 4-8 animals). Shown in (d) are total numbers of clones in each of the dentate gyrus examined for both control and  $\gamma_2$  cKO adult mice under basal conditions. Each dot represents data from one dentate gyrus. Red lines represent mean values. **e-f**, Characterization of the efficacy of  $\gamma_2$  deletion in the clonal analysis. Shown in (e) are sample confocal images of immunostaining of GFP, GFAP and  $\gamma_2$ . Scale bar: 10  $\mu$ m. Shown in (f) are representative whole-cell recording traces of GABAergic synaptic currents from GFP<sup>+</sup> neurons before and after application of diazepam (1  $\mu$ M) and BMI (50  $\mu$ M) at 30 dpi in acute slices prepared from induced control and cKO mice. Numbers indicate number of clones with the phenotype among all recorded clones. **g**, A schematic diagram of experimental scheme to analyze the effect of systemic diazepam application in clonal analysis (related to Fig. 2e & 3d). After a single dose of tamoxifen injection (62 mg/kg body weight), vehicle or diazepam (5 mg/kg body weight) was *i.p.* injected once daily for 6 days, and animals were processed for immunohistology one day later for clonal analysis.



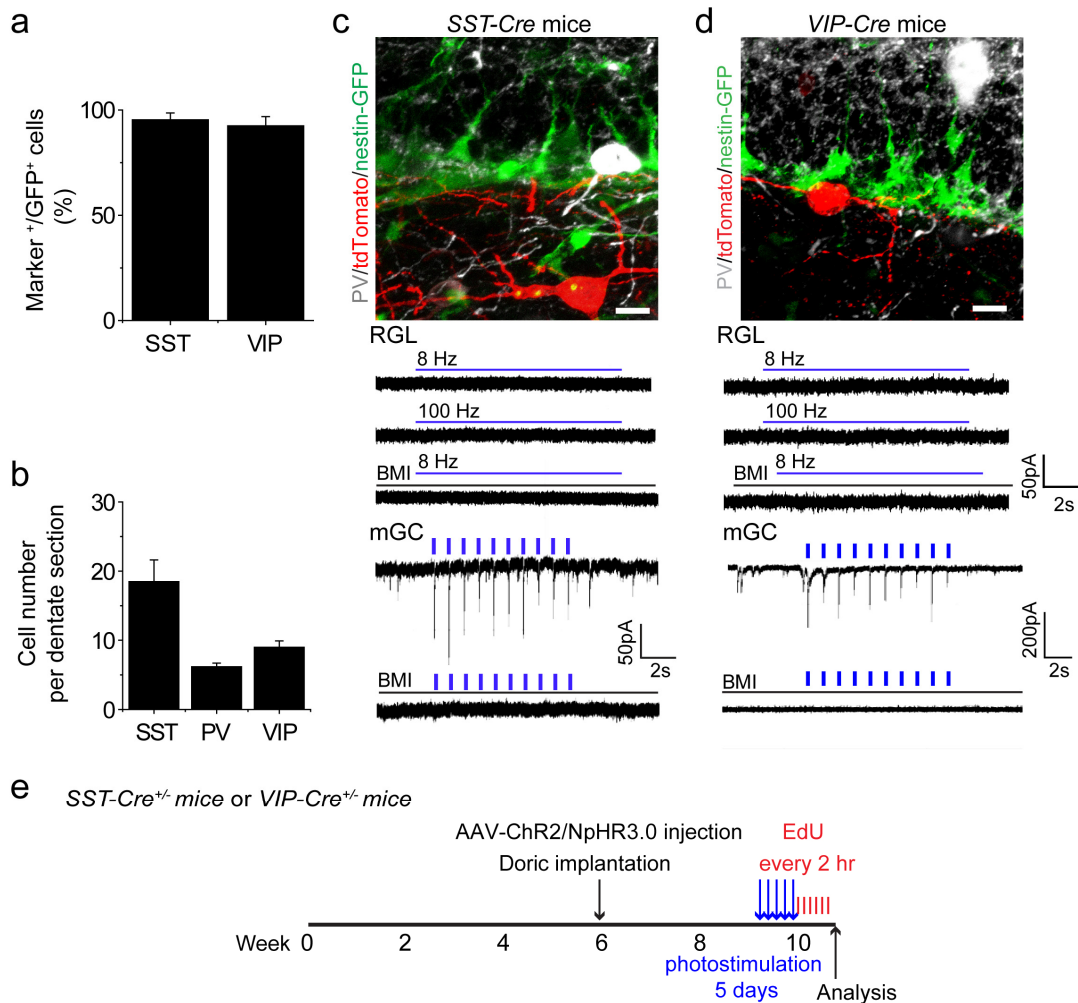
**Supplementary Figure 3. Clonal analysis of  $\gamma_2$  function in regulating RGLs at 30 dpi in the adult dentate gyrus.** **a-c**, Sample confocal images of clones exhibiting active self-renewal. Shown in **(a)** is a GFP<sup>+</sup> clone consisting of a nestin<sup>+</sup>GFAP<sup>+</sup> RGL (1) and a nestin<sup>-</sup>GFAP<sup>+</sup> bushy astrocyte (2), indicating self-renewal and unipotential astrogenic differentiation. Shown in **(b)** is a GFP<sup>+</sup> clone consisting of a GFAP<sup>+</sup>MCM2<sup>-</sup> RGL (1) and GFAP<sup>+</sup>MCM2<sup>+</sup> IPCs (2), indicating self-renewal and unipotential neurogenic differentiation. Shown in **(c)** is a GFP<sup>+</sup> clone consisting of a GFAP<sup>+</sup> RGL (1), a GFAP<sup>+</sup> bushy astrocyte (2), and a cluster of cells of the neuronal lineage (3), some of which expressed immature neuronal marker PSA-NCAM, indicating self-renewal and multi-lineage differentiation. Scale bars: 10  $\mu$ m. **d-e**, Quantitative comparison of the frequency of different clone types between control (n = 7) and  $\gamma_2$  cKO (n = 6) at 30 dpi. Shown are pie charts of the relative frequency of quiescence/activation (**d**) and the summary of the frequency of different types among all RGL-containing GFP<sup>+</sup> clones (**e**).



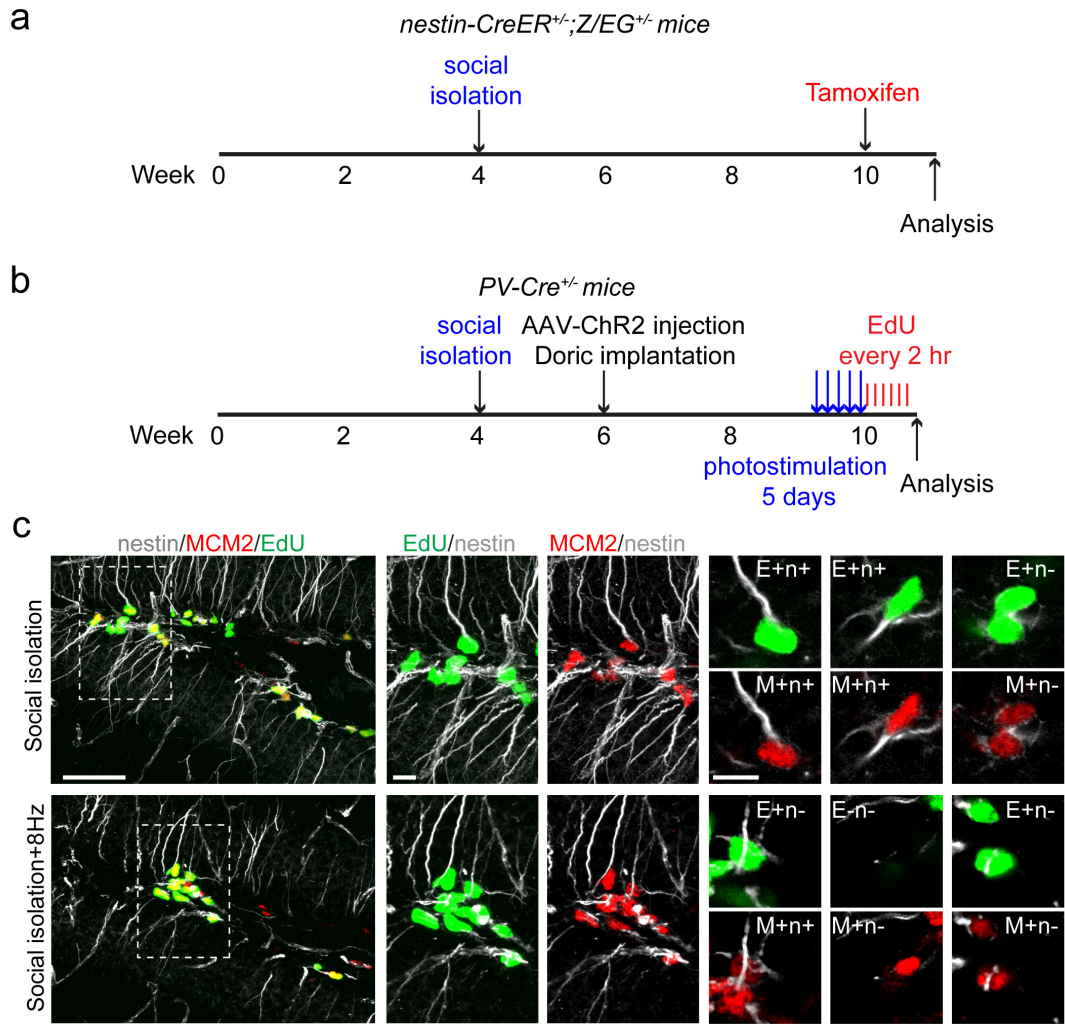
**Supplementary Figure 4. Characterization of optogenetic tools for manipulating PV<sup>+</sup> interneuron activity in the adult dentate gyrus.** **a**, A schematic diagram of AAV constructs used for Cre-dependent expression of ChR2 or eNpHR in PV<sup>+</sup> interneurons using adult *PV-Cre* mice. **b**, Sample confocal images of expression of ChR2-YFP or eNpHR-YFP and PV immunostaining 2 weeks after AAV-DIO-ChR2-YFP or AAV-DIO-NpHR-YFP injection into the dentate gyrus of adult *PV-Cre* transgenic mice. Scale bars: 50  $\mu$ m. **c**, Quantification of the specificity of AAV-mediated expression in PV<sup>+</sup> interneurons in the adult dentate gyrus. Shown are summaries for both YFP and mCherry tagged opsin vectors ( $n = 5-10$  animals). **d-e**, Effective photo-activation or suppression of PV<sup>+</sup> interneurons in slices acutely prepared from AAV injected animals. Shown are sample traces of whole-cell recordings of YFP<sup>+</sup>PV<sup>+</sup> interneurons under both current-clamp (upper traces) and voltage-clamp modes (lower traces) upon photo-activation (**d**; 472 nm blue light at 8 Hz, 5 ms) or upon photo-suppression (**e**; 593 nm yellow light; constant). Also shown is the quantification of the efficiency of photo-induced suppression. A current pulse of 100 pA current was delivered to YFP<sup>+</sup>PV<sup>+</sup> interneurons, and the number of action potentials with or without yellow light on was quantified. Values represent mean  $\pm$  s.e.m. ( $n = 6$  cells; \*:  $P < 0.05$ ; student t-test).



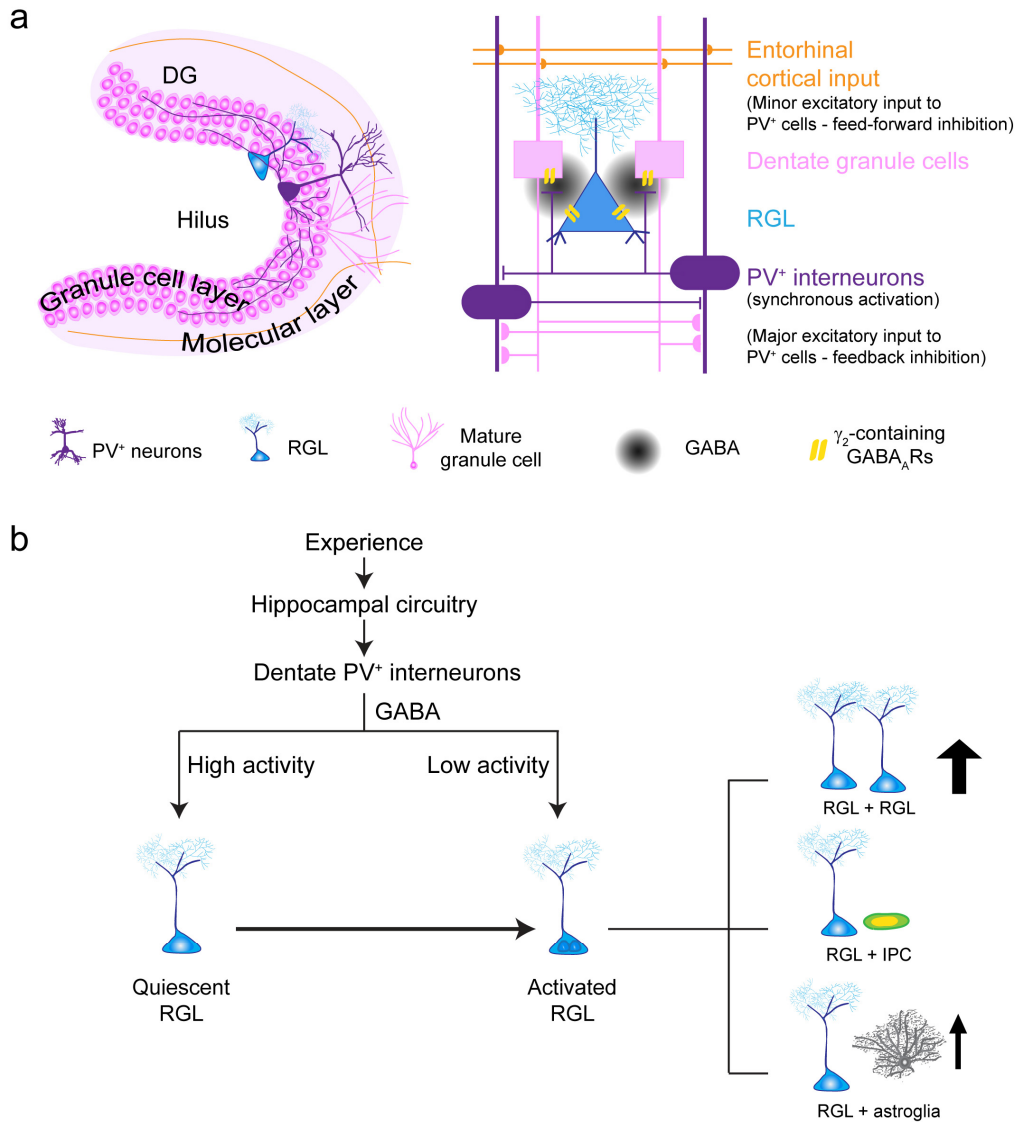
**Supplementary Figure 5. Analysis of effects of *in vivo* manipulation of PV<sup>+</sup> interneuron activity on RGL activation in the adult dentate gyrus of PV-Cre mice. a,** A schematic diagram of analysis of RGL activation at the population level upon photo-activation or photo-suppression of PV<sup>+</sup> interneurons in the dentate gyrus of adult PV-Cre mice (related to **Fig. 4d**). For photo-activation of PV<sup>+</sup> interneurons, blue light flashes (472 nm; 5 ms at 8 Hz) were delivered every 5 minutes for 30s/per trial, 8 hrs per day for 5 days. For photo-suppression of PV<sup>+</sup> interneurons, continuous yellow light (593 nm; constant) was delivered 8 hrs per day for 5 days. On the 5<sup>th</sup> day, animals were *i.p.* injected with EdU (41.1 mg/kg body weight) 6 times 2 hrs apart and processed for immunohistology 2 hrs after the last EdU injection. **b,** Sample confocal images of immunostaining of nestin (white), MCM2 (red), EdU (green) and DAPI (blue). Yellow arrows point to double labelled RGLs (nestin<sup>+</sup>MCM2<sup>+</sup> or nestin<sup>+</sup>EdU<sup>+</sup> cells with radial process) and orange arrows point to EdU<sup>-</sup> or MCM2<sup>-</sup> nestin<sup>+</sup> cells. Scale bars: 50 μm and 10 μm (insert).



**Supplementary Figure 6. Characterization of optogenetic tools for manipulating SST<sup>+</sup> and VIP<sup>+</sup> interneuron activity in the adult dentate gyrus.** **a-b**, Characterization of opsin expression in different animal models. Shown in **(a)** is a quantification of immunohistological analysis of the specificity of ChR2 expression in SST<sup>+</sup> and VIP<sup>+</sup> interneurons after AAV-DIO-ChR2-YFP injection into the dentate gyrus of adult SST-Cre or VIP-Cre mice, respectively. Shown in **(b)** is a summary of quantification of numbers of ChR2-YFP labelled neurons after AAV-DIO-ChR2-YFP injection into the dentate gyrus of adult PV-Cre, SST-Cre or VIP-Cre mice. Values represent mean  $\pm$  s.e.m. ( $n = 3-10$  animals). **c**, Lack of responses of RGLs to SST<sup>+</sup> interneuron activation. Shown at the top is a sample confocal image of nestin-GFP, ChR2-tdTomato, and PV immunostaining in the dentate gyrus of adult SST-Cre<sup>+/+</sup>nestin-GFP<sup>+/+</sup> mice after AAV-DIO-ChR2-tdTomato injection. Scale bar: 10  $\mu$ m. Shown at the bottom are sample whole-cell voltage-clamp recording traces from a GFP<sup>+</sup> RGL upon light stimulation of SST<sup>+</sup> neurons (472 nm blue light at 8 or 100 Hz) and from a mature dentate granule cell (mGC; at 1 Hz) upon light stimulation of SST<sup>+</sup> interneurons in the acute slices before and after bicuculline treatment (BMI; 50  $\mu$ M). **d**, Lack of responses of RGLs to VIP<sup>+</sup> interneuron activation. Similar as in **(c)**, except adult VIP-Cre<sup>+/+</sup>nestin-GFP<sup>+/+</sup> mice were used. **e**, A schematic diagram of analysis of RGL activation at the population level upon photo-activation or photo-suppression of SST<sup>+</sup> or VIP<sup>+</sup> interneurons in the adult dentate gyrus. The same paradigm as in Supplementary Fig. 5a was used.



**Supplementary Figure 7. Experimental schemes involving social isolation.** **a**, A schematic diagram of clonal analysis of RGLs upon social isolation (related to **Fig. 5a-b**). **b**, A schematic diagram of analysis of RGL activation at the population level upon photo-stimulation of PV<sup>+</sup> interneurons in the adult dentate gyrus following social isolation (relate to **Fig. 5c**). Same paradigm of photo-stimulation and EdU injections as in Supplementary Fig. 5a was used. **c**, Sample confocal images of immunostaining for nestin (white; n), MCM2 (red; M), and EdU (green; E). Scale bars: 50  $\mu$ m (left column) and 10  $\mu$ m for inserts (five right columns).



**Supplementary Figure 8. A summary model of regulation of quiescent adult neural stem cells *in vivo* in response to neuronal activity and experience, involving GABA release from local PV<sup>+</sup> interneurons and  $\gamma_2$ -containing GABA<sub>A</sub>R-mediated signalling in RGLs in the adult dentate gyrus. **a**, Schematic diagrams of local dentate gyrus circuitry. PV<sup>+</sup> interneurons receive excitatory inputs from both dentate granule cells (90%) and entorhinal cortical inputs (5%). PV<sup>+</sup> interneurons are also coupled with each other and fire synchronously. PV<sup>+</sup> interneurons innervate dentate granule cell layer and spillover of GABA from synaptic junctions regulates nearby radial glia-like neural stem cells (RGLs). **b**, A model of neuronal activity-dependent regulation of radial glia-like neural stem cell fate decision in the adult dentate gyrus. High levels of PV<sup>+</sup> interneuron activity promote quiescence of neural stem cells, whereas low levels of PV<sup>+</sup> interneuron activity promote activation of neural stem cells and symmetric cell division.**



## Supplementary Movies

**Supplementary Movie 1. Close association between GFP<sup>+</sup> RGLs and GAD67<sup>+</sup> terminals of PV<sup>+</sup> interneurons in the adult dentate gyrus.** Shown is a surface-rendered reconstruction of confocal images of the dentate gyrus (90 x 90 x 30  $\mu\text{m}$ ) of adult *nestin-GFP* mice for immunostaining of nestin-GFP (green), PV<sup>+</sup>GAD67<sup>+</sup> (blue) and PV<sup>+</sup>GAD67<sup>+</sup> (red).

**Supplementary Movie 2. Lack of interaction between SST<sup>+</sup> interneurons and RGLs in the adult dentate gyrus.** Shown is a surface-rendered reconstruction of confocal images of PV immunostaining (white), ChR2-tdTomato expression by SST<sup>+</sup> interneurons (red) and nestin-GFP expression in RGLs (green). DIO-ChR2-mCherry AAV was injected into *SST-Cre<sup>+/+</sup>;nestin-GFP<sup>+/+</sup>* mice and sections were processed for immunostaining two weeks later. Note that SST<sup>+</sup> interneurons extend neuronal processes in the subgranular zone and hilus region, whereas PV<sup>+</sup> interneurons exhibit elaborated processes in the subgranular zone and granule cell layer in the adult dentate gyrus.

**Supplementary Movie 3. A single PV<sup>+</sup> interneuron has the capacity to regulate a large number of RGLs in the adult dentate gyrus.** Shown is a surface reconstruction of serial confocal images of a single PV<sup>+</sup> interneuron in the adult dentate gyrus. A small volume of DIO-mCherry AAV was injected into *PV-Cre<sup>+/+</sup>;nestin-GFP<sup>+/+</sup>* mice and a region with a single PV<sup>+</sup> neuron was confocal imaged for mCherry (PV<sup>+</sup> neuron) and GFP (nestin-expressing cells) 2 weeks after injection and reconstructed from 8 serial coronal sections (50  $\mu\text{m}$  in thickness per section). An estimated 200+ GFP<sup>+</sup> RGLs were covered by a single PV<sup>+</sup> interneuron in the adult dentate gyrus.